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TRANSMISSION OF RADISH-MOSAIC VIRUS BY APHIDS¹

HENRY H. P. SEVERIN² and C. M. TOMPKINS³

INTRODUCTION

A MOSAIC DISEASE of the cultivated radish (Raphanus sativus) is common in the vegetable gardens of the San Francisco Bay districts. The disease also occurs on wild radish, which probably serves as a reservoir of the virus.

Tompkins (1939)⁴ reviewed the literature on mosaic diseases of Raphanus species. He also published the results of studies on host range and properties of the radish-mosaic virus. The virus was found to be readily transmitted by mechanical inoculation. Numerous unsuccessful attempts were made to transmit the virus by means of the cabbage aphid, Brevicoryne brassicae (L.), turnip or false cabbage aphid, Rhopalosiphum pseudobrassicae (Davis), and the green peach aphid, Myzus persicae (Sulzer).

Magistad (1938) listed daikon (Raphanus sp.) mosaic in the Hawaiian Islands.

Parris (1940) listed a mosaic on Raphanus sp. in Hawaii.

Dale (1948) recorded the occurrence of a mosaic disease on three crucifers in Trinidad, British West Indies, one of which was an eastern variety of radish, known locally as moorei and which he suggested may be Raphanus sativus var. hortensis. He found by inoculation tests that the radish (R. sativus) varieties French Breakfast and Burpee's Red Giant appear to be immune to the virus described in his paper. The virus was transmitted by Rhopalosiphum pseudobrassicae (Davis).

The following is a report of a study undertaken on aphid transmission of the radish-mosaic virus described by Tompkins (1939). Tests were made with aphid species that breed on White Icicle radish under natural conditions and with species that have not been reared on this host plant. A comparison was made between transmission of the virus by unfasted infective aphids, and transmission by previously noninfective fasted aphids after short infection-feeding intervals. Other aspects investigated include a comparison of virus transmission, by lots of varying numbers of aphids, with mechanical inoculation; and the retention of the virus.

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^{*} See "Literature Cited" for citations referred to in the text by author and date.

MATERIALS AND METHODS

The virus was obtained from a naturally infected white radish plant grown near San Pablo, Contra Costa County. It was maintained by repeated mechanical inoculation and aphid transmission of the virus to healthy White Icicle radish seedlings. The carborundum method (Rawlins and Tompkins, 1936) was used in mechanical inoculation. White Icicle radish seedlings grown from seeds were used in all experiments reported in this paper.

Colonies of noninfective aphid species have been reared in the greenhouse for a period of 20 years. The methods of collecting, rearing, and transferring aphids, and of producing noninfective aphids, have been described in previous papers (Severin and Freitag, 1938). Another method of obtaining noninfective aphids has been reported by Severin and Tompkins, 1948.

SYMPTOMATOLOGY

The first symptom of radish mosaic on plants infected with the virus by aphids or mechanical inoculation usually is a clearing of the veins and veinlets (plate 1, A), first appearing on the outer or intermediate leaves and spreading to the youngest leaves, followed by yellow veinbanding (plate 1, B). The yellow veinbanding of the veinlets then encloses green, slightly raised areas (plate 2, A, B) which later turn vellow (plate 2, C). Sometimes, however, the first symptom consists of small, circular, chlorotic areas (plate 3, A) between and adjacent to the veins. Fusion of these chlorotic areas later occurs (plate 3, B). Within a few days the chlorotic areas become numerous (plate 3, C) and soon replace the normal green tissue with a coarse mottle (plate 3, D) followed by necrosis (plate 4, A). Sometimes vellow veinbanding of the cleared veinlets and small, circular, chlorotic areas occur on the same leaf (plate 4, B). In the later stage of the disease variable mosaic patterns occur, consisting of chlorotic veinbanding (plate 1, C, plate 6, C), interveinal chlorosis (plate 1, C, plate 5, A, B), marginal chlorosis (plate 1, D), green veinbanding (plate 6, A, B), occasionally slightly raised dark-green or blisterlike areas (plate 2, D, plate 6, D). The chlorosis gradually spreads until the leaf becomes yellow (plate 5, C, D). In the advanced stage of the disease the youngest leaves are dwarfed and curled (plate 7).

TRANSMISSION OF VIRUS

By Vectors that Breed on Radish under Natural Conditions. Four species of aphids which multiply on radish under natural conditions are vectors of the virus. These are:

Cabbage aphid, Brevicoryne brassicae (L.) Turnip or false cabbage aphid, Rhopalosiphum pseudobrassicae (Davis) Ornate aphid, Myzus ornatus Laing Green peach aphid, Myzus persicae (Sulzer)

By Vectors that Do Not Breed on Radish. The following eight species of aphids which do not multiply on radish in the greenhouse are vectors of the virus:

Celery aphid, Aphis apii Theobald Rusty-banded aphid, Aphis ferruginea-striata Essig Cotton or melon aphid, Aphis gossypii Glover Bean or dock aphid, Aphis rumicis L. Yellow willow aphid, Cavariella aegopodii (Fabricius) Pea aphid, Macrosiphum pisi (Kaltenbach) Lily aphid, Myzus circumflexus (Buckton) Foxglove aphid, Myzus solani (Kaltenbach)

By Unstarved Single Infective Aphids that Multiply on Radish. Four species of unstarved, infective, single aphids that breed on radish outdoors were tested as vectors of the virus. These, the cabbage, false cabbage or turnip, ornate, and green peach aphids were reared on mosaic-infected radish plants,

TABLE 1

TRANSMISSION OF RADISH-MOSAIC VIRUS BY SPECIES OF UNSTARVED, SINGLE, WINGLESS, INFECTIVE APHIDS FROM DISEASED TO HEALTHY PLANTS

Common and scientific names of aphids	Plants inoculated	Plants infected	Per cent infected
Celery aphid, Aphis apii*	50	1	2.0
Rusty banded aphid, Aphis ferruginea-striata		1	2.0
Cotton or melon aphid, Aphis gossypii	25	4	8.0
Bean or dock aphid, A phis rumicis.	100	2	2.0
Cabbage aphid, Brevicoryne brassicae	25	9	36.0
Yellow willow aphid, Cavariella aegopodii	25	6	24.0
Pea aphid, Macrosiphum pisi	25	11	22.0
Lily aphid, Myzus circumflexus	50	6	24.0
Ornate aphid, Myzus ornatus.		10	10.0
Green peach aphid, Myzus persicae	100	12	48.0
Foxglove aphid, Myzus solani	25	5	20.0
Honeysuckle aphid, Rhopalosiphum conii	25	1	2.0
Turnip aphid, Rhopalosiphum pseudobrassicae	25	3	12.0

* According to E. O. Essig (personal interview) A phis april Theobald may be identical with A. helianthi Monell.

and then each aphid was fed singly on a healthy radish seedling. Table 1 shows that the cabbage aphid infected 36 per cent, the turnip aphid 12 per cent, the ornate aphid 10 per cent, and the green peach aphid 48 per cent of the 25 to 100 plants inoculated with each species.

By Previously Noninfective, Starved, Single Aphids in Short Infectionfeeding Periods. Previously noninfective, starved, wingless aphids were fasted two to three hours in a phial, and then were fed singly on a mosaicinfected radish plant for one-half, one, two, five, or ten minutes, and then on a healthy radish seedling. A comparison of the results obtained in tables 1 and 2 shows that higher percentages of transmission of the virus were obtained with starved single aphids in short infection-feeding periods than with unstarved single aphids, with the exception of the bean or dock, cabbage, yellow willow, and pea aphids. Table 2 shows that the bean or dock and honeysuckle aphids failed to transmit the virus.

COMPARATIVE EFFICIENCY OF MECHANICAL INOCULATION AND APHID TRANSMISSION OF VIRUS

A comparison was made of mechanical inoculation and aphid transmission of the virus. In each test, a single species of infective wingless aphids was transferred, in lots of 10 and 20, from infected to healthy plants. Thirteen TABLE 2

TRANSMISSION OF RADISH-MOSAIC VIRUS BY SPECIES OF PREVIOUSLY STARVED, SINGLE, WINGLESS APHIDS AFTER SHORT INFECTION-FEEDING PERIODS ON VIRUS SOURCE

					Feeding	Feeding time, minutes, on diseased leaves	nutes, oi	ı disease	d leaves				
Common and scientific names of aphids	0.5	2	1		3		5		1	0	Totals	als	
	Plants inocu- lated	Plants in- fected	Plants inocu- lated	Plants in- fected	Plants inocu- lated	Plants in- fected	Plants inocu- lated	Plants in- fected	Plants inocu- lated	Plants in- fected	Plants inocu- lated	Plants in- fected	Per cent in- fected
Celery aphid, A phis apii	10	5	10	3	10	÷	10	1	10	5	50	11	22.0
Rusty-banded aphid, Aphis ferruginea-striata	5	0	5	0	ç	0	ŝ	0	ñ	1	25	1	4.0
Cotton or melon aphid, A phis gossy pii	5	-	ŝ	0	S	67	ŝ	-	ŝ	1	25	5	20.0
Bean or dock aphid, Aphis rumicis	£.	:	ñ	:	5	:	5	:	ŝ	:	50	0	0.0
Cabbage aphid, Brevicoryne brassicae	5	0	2	67	ç	0	ŝ	-	ŝ	0	25	en	12.0
Yellow willow aphid, Cavariella aegopodii	•••	:	ŝ	:	5	:	'n	:	ŝ	:	25	5	20.0
Pea aphid, Macrosiphum pisi.	'n	0	ō	1	5	0	ñ		ŝ	0	25	61	8.0
Lily aphid, Myzus circumflexus	ñ	5	2	5	5	ŝ	ŝ	Ч	5	4	25	12	48.0
Ornate aphid, Myzus ornatus	'n	67	5	5	5	0	ŝ	-	ŝ	63	25	7	28.0
Green peach aphid, Myzus persicae	10	ŝ	10	5	10	e	10	ŝ	10	1	25	12	48.0
Foxglove aphid, Myzus solani	مر	ŝ	ũ	2	5	61	ŝ	ŝ	5	1	25	Ξ	44.0
Honeysuckle aphid, Rhopalosiphum conii	ŝ	0	2	0	5	0	ñ	0	5	0	25	0	0.0
Turnip aphid, Rhopalosiphum pseudobrassicae	ŝ	4	5	en	ŝ	ŝ	ŝ	1	ŝ	ŝ	25	14	56.0
		_	-	-								-	

* Aphids crawled about on leaf, some fed, feeding time y_2 to 10 minutes.

TABLE 3	
COMPARISON OF TRANSMISSION OF RADISH-MOSAIC VIRUS BY	Y
MECHANICAL INOCULATION WITH SPECIES OF	
APHID, 20 IN EACH LOT	

Mech	anical inoc	ulation	Aph	Aphid transmission							
Plants inocu- lated	Plants infected	Per cent infected	Common and scientific names of aphids	Number of aphids	Plants inocu- lated	Plants infected	Per cent infected				
10	3	30.0	Celery aphid, Aphis apii	10	10	5	50.0				
10	10	100.0		20	10	6	60.0				
10	10	100.0	Rusty-banded aphid, Aphis	10	10	4	40.0				
10	0	0.0	ferruginea-striata	20	10	1	10.0				
10	9	90.0	Cotton or melon aphid, Aphis	10	10	6	60.0				
10	3	30.0	gossy pii	20	10	10	100.0				
60	12	20.0	Bean or dock aphid, Aphis	10	60	0	0.0				
20	7	70.0	rumicis	20	10	0	0.0				
10	9	90.0	Cabbage aphid, Brevicoryne	10	10	10	100.0				
25	20	80.0	brassicae	20	25	18	72.0				
10	8	80.0	Yellow willow aphid, Cavariella	10	10	7	70.0				
10	7	70.0	aegopodii	20	10	8	80.0				
10	10	100.0	Pea aphid, Macrosiphum pisi	20	10	9	90.0				
10	10	100.0		10	10	10	100.0				
10	5	50.0	Lily aphid, Myzus circumflexus	10	10	6	60.0				
10	8	80.0		20	10	7	70.0				
10	8	80.0	Ornate aphid, Myzus ornatus	10	10	9	90.0				
20	4	20.0		20	10	2	20.0				
10	10	100.0	Green peach aphid, Myzus	10	10	5	50.0				
10	8	80.0	persicae	20	10	1	10.0				
10	9	90.0	Foxglove aphid, Myzus solani	10	10	2	20.0				
10	4	40.0		20	20	6	30.0				
20	16	80.0	Honeysuckle aphid,	10	20	0	0.0				
20	15	75.0	Rhopalosiphum conii	20	20	0	0.0				
20	14	70.0	Turnip aphid, Rhopalosiphum	10	20	8	40.0				
10	4	40.0	pseudobrassicae	20	10	8	80.0				

species were so tested. Also, the virus from the same infected plants upon which the aphids had fed was mechanically inoculated in healthy plants. The results obtained are given in table 3.

Of 13 species of aphids tested, the pea aphid was the most efficient vector. The bean or dock and honeysuckle aphids failed to transmit the virus. A comparison of the transmission of the virus by lots of 10 aphids with lots of 20 aphids (excepting the two species which failed to infect any of the plants) shows that in the ten-aphid lots a total of 72 of 120 plants, or 56 per cent, were infected; while in the 20 aphid lots 76 of 135 plants, or 60 per cent, were infected.

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Of the 255 plants mechanically inoculated, 173 plants or 68 per cent were infected, omitting the results of mechanical inoculation with the two species which failed to transmit the virus. It is evident that the 11 species of aphids were less efficient than mechanical inoculation in transmitting the virus.

RETENTION OF VIRUS

By Lots of 20 Aphids in Daily Transfers. Retention of the virus was determined for four species of aphids reared on infected radish plants. Lots of 20 aphids of each species were transferred daily for three days to successive healthy plants. As table 4 shows, each of the four species transmitted the virus from diseased to healthy plants during the first day but none of the lots produced infections during the second and third days.

TABLE 4 RETENTION OF RADISH-MOSAIC VIRUS BY FOUR SPECIES OF APHIDS, 20 IN EACH LOT

	Number	First	t d ay	Secon	d day	Thir	d day
Aphid species	Number of lots	Plants inocu- lated	Plants in- fected	Plants inocu- lated	Plants in- fected	Plants inocu- lated	Plants in- fected
Cabbage aphid, Brevicoryne brassicae Turnip aphid, Rhopalosiphum	5	5	5	5	0	5	0
pseudobrassicae	5	5	5	5	0	5	0
Ornate aphid, Myzus ornatus	5	5	4	5	0	5	0
Green peach aphid, Myzus persicae	5	5	3	5	0	5	0

By Lots of 20 Aphids with Hourly Transfers. An experiment was conducted to determine more precisely how long aphids of four species would retain the virus. Lots of 20 infective, wingless aphids reared on infected radish plants were transferred hourly to six successive healthy plants.

As table 5 shows, one lot of cabbage aphids infected three successive plants during the first three hours, two lots during the first two hours, and two lots during the first hour only. One lot of turnip aphids produced infections in three successive plants during the first three hours, and four lots during the first two hours. Two lots of ornate aphids infected three successive plants during the first and second hours, and three lots during the first hour only. Three lots of green peach aphids caused infections during the first hour only.

Conclusion. The virus is of the nonpersistent type (Watson, 1939, Watson and Roberts, 1940) in the aphid vectors, and was retained from one to three hours. According to Tompkins (1939) the virus remained active *in vitro* for 14 days at 22° C, but was inactivated after 16 days.

TABLE 5

RETENTION OF RADISH-MOSAIC VIRUS BY FOUR SPECIES OF APHIDS TRANSFERRED HOURLY TO SIX SUCCESSIVE HEALTHY PLANTS

Aphid species and lot no.	Number of aphids	Results* on successive plants, with hourly transfers						Last infection produced
	on first plant	1st	2d	3d	4th	5th	6th	by aphids, hour
Cabbage aphid, Brevicoryne brassicae								
Lot 1	20	+	+	+	-	-	-	3d
Lot 2	20	+	+		-	-	-	2d
Lot 3	20	+	+	-	-	-	-	2d
Lot 4	20	+	-	-	-	—		1st
Lot 5	20	+	-	-	-	-	- 1	lst
Turnip aphid, Rhopalosiphum pseudobrassicae								
Lot 1	20	+	+	+	-	-		. 3d
Lot 2	20	+	+		- 1		-	2d
Lot 3	20	+	+	-		-	-	2d
Lot 4	20	+	+	-	-	_	-	2d
Lot 5	20	+	+		-	-	-	2d
Ornate aphid, Myzus ornatus								
Lot 1	20	+	+	-	-	-	-	2d
Lot 2	20	+	+		-	-	-	2d
Lot 3	20	+	-	-	-	-	-	1st
Lot 4	20	+	-	-	-	-	-	1st
Lot 5	20	+	-	-	-	_	-	1st
Green peach aphid, Myzus persicae								
Lot 1	20	+	_	-	-		-	1st
Lot 2	20	+	_	-	-	-	-	1st
Lot 3	20	+	-	-	-	-	-	1st
Total +		18	10	2	0	0	0	
Total –		0	8	16	18	18	18	

* The plus sign (+) indicates the production of the disease, and the minus (-) shows that no disease resulted.

SUMMARY

The symptoms of radish mosaic are described on experimentally infected plants. Four species of aphids which multiply on radish under natural conditions are vectors of the virus. These are:

Cabbage aphid, Brevicoryne brassicae (L.) Turnip or false cabbage aphid, Rhopalosiphum pseudobrassicae (Davis) Ornate aphid, Myzus ornatus Laing Green peach aphid, Myzus persicae (Sulzer)

The following eight species of aphids which do not multiply on radish in the greenhouse are vectors of the virus:

Celery aphid, Aphis apii Theobald Rusty-banded aphid, Aphis ferruginea-striata Essig Cotton or melon aphid, Aphis gossypii Glover Bean or dock aphid, Aphis rumicis L. Yellow willow aphid, Cavariella aegopodii (Fabricius) Pea aphid, Macrosiphum pisi (Kaltenbach) Lily aphid, Myzus circumflexus (Buckton) Foxglove aphid, Myzus solani (Kaltenbach) Hilgardia

Higher percentages of transmission of the virus were obtained with previously noninfective, starved aphids in short infection-feeding intervals than with unstarved, infective aphids, with the exception of the bean or dock, cabbage, yellow willow, and pea aphids.

Mechanical inoculation (68 per cent) was more efficient than transmission of the virus by 11 species of aphids (54 per cent).

In tests on retention of the virus with four species of aphids, 20 in each lot, all species transmitted the virus during the first day, but none of the lots produced infections during the second and third days.

In tests with hourly transfers, transmissions occurred within the first three hours after the aphids had fed on a mosaic-infected plant.

The radish-mosaic virus is a nonpersistent type in the aphid vectors.

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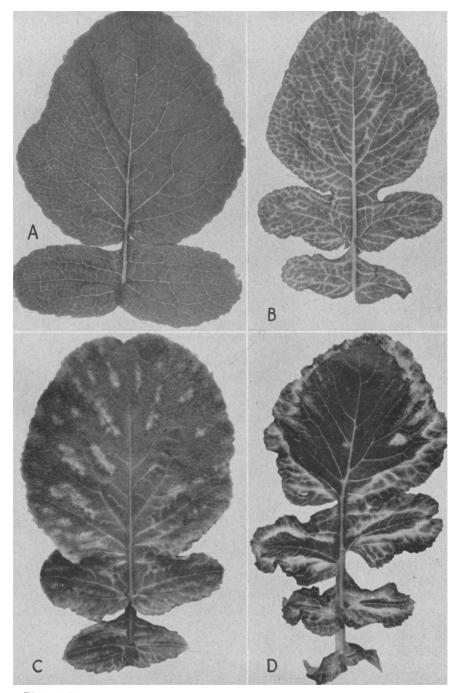


Plate 1. Symptoms of mosaic on leaves of White Icicle radish (*Baphanus sativus*): A, cleared veins and veinlets; B, yellow veinbanding of clear veins and veinlets; C, chlorotic veinbanding; D, marginal chlorosis.

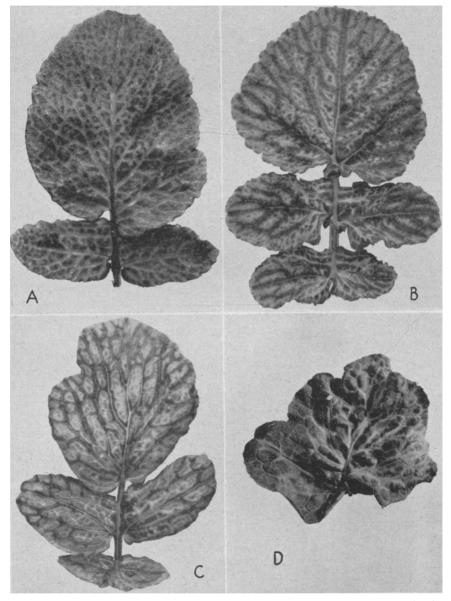


Plate 2. Symptoms of mosaic on leaves of White Icicle radish (*Raphanus sativus*): A, B, C, successive stages of yellow veinbanding enclosing slight raised green areas; D, blisterlike elevations.

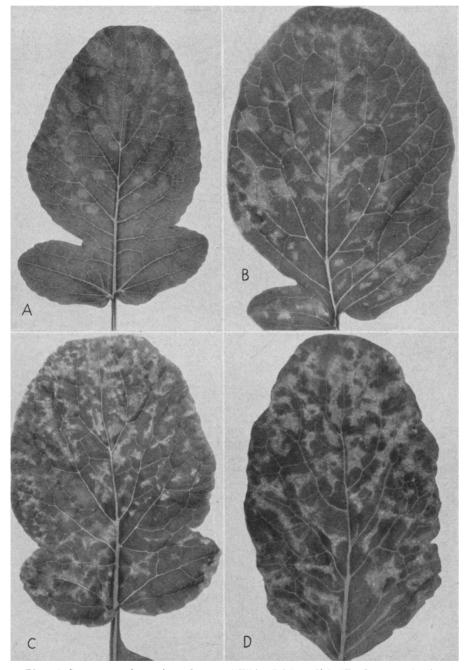


Plate 3. Symptoms of mosaic on leaves of White Icicle radish (*Raphanus sativus*): A, circular, chlorotic areas between and adjacent to the veins; B, fusion of chlorotic areas; C, D, numerous fused chlorotic areas, replacing the normal green tissue with a coarse mottle.

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[SEVERIN-TOMPKINS] PLATE 4

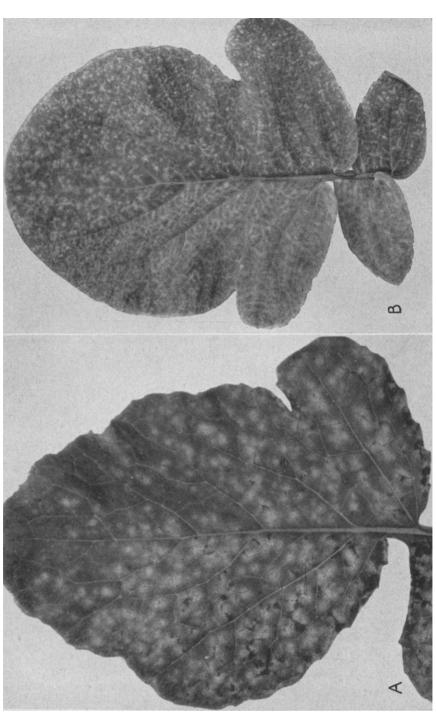


Plate 4. Symptoms of mosaic on leaves of White Icicle radish (Raphanus satious): A, necrosis of chlorotic areas; B, yellow veinbanding of cleared veins and veinlets and small, circular, chlorotic areas.

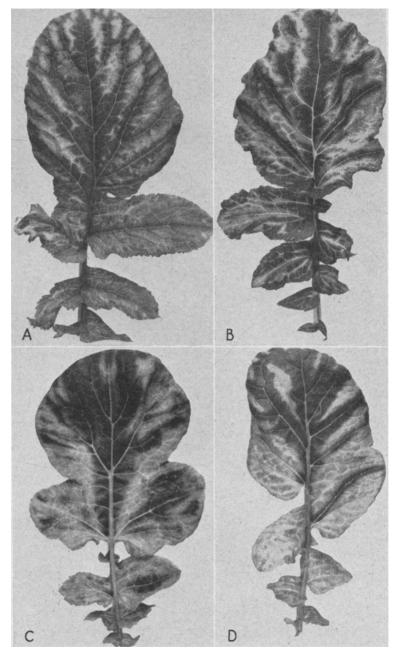


Plate 5. Symptoms of mosaic on leaves of White Icicle radish (*Raphanus sativus*): A, B, interveinal chlorosis; C, D, chlorosis spreading until leaves become yellow.

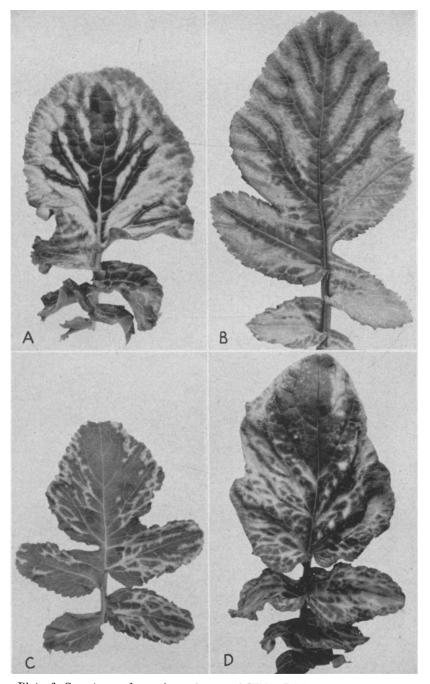


Plate 6. Symptoms of mosaic on leaves of White Icicle radish (*Raphanus sativus*): A, B, green veinbanding; C, chlorotic veinbanding; D, blisterlike elevations.



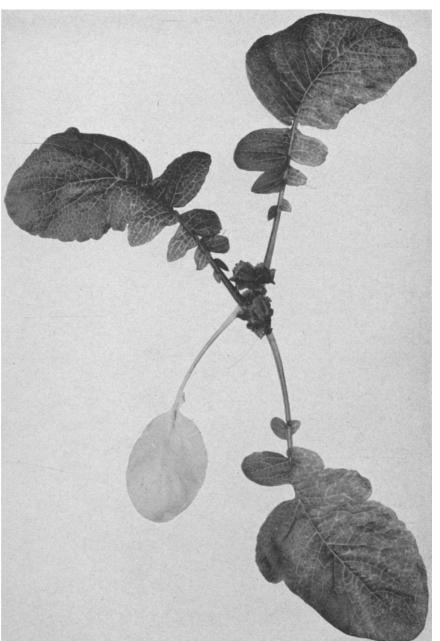


Plate 7. Symptoms of mosaic on leaves of White Icicle radish (Raphanus sativus) showing cleared veins and veinlets on older leaves, and dwarfed, curled, youngest leaves.

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